Assessment of CASP gene polymorphisms in periodontal disease

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ABSTRACT. Caspases (CASP) are intracellular proteases that play roles as mediators of apoptosis. Activation of caspase 3 is enhanced in chronic periodontitis. Thus, we hypothesized that single nucleotide polymorphisms (SNPs) of CASP genes might be associated with this condition in the Korean population. To investigate whether such polymorphisms might be involved in the development of periodontal disease, 51 patients and 33 control subjects were assessed. A total of 201 CASP gene SNPs were analyzed with genotypes being determined using and Axiom™ genome-wide human assay. SNPStats and SPSS 18.0 were used for the analysis of genetic data and logistic regression models were utilized to evaluate odds ratios, 95% confidence intervals, and P values. Of the 201 SNPs, only three (rs12108497, rs4647602, and rs113420705, all in the CASP3 gene) were significantly associated with chronic periodontitis (P < 0.05). The minor allele frequencies of these SNPs were higher in the patient group than in the control group. In addition, the TC and GT haplotypes
formed by rs4647602 and rs113420705 were found to be associated with chronic this disease (TC haplotype, $P = 0.0039$; GT haplotype, $P = 0.002$). These results suggest that CASP3 gene polymorphisms may be associated with susceptibility to periodontal disease in the Korean population.

**Key words:** Association study; Periodontal disease; CASP genes; Polymorphism

**INTRODUCTION**

Tooth loss is highly associated with quality of life (Gerritsen et al., 2010). Periodontitis, a major cause of tooth loss, is common among adults in all regions and is found at a rate of 5-15% in most populations (Burt et al., 1990; Petersen, 2003). Between 2009 and 2012, 46% of US adults, representing 64.7 million people, suffered periodontitis, with 8.9% constituting severe cases (Eke et al., 2015). The occurrence of periodontitis is associated with sex/gender, race/ethnicity/skin color, education/schooling, socioeconomic classification/status/position, and income (Oppermann et al., 2015). In industrialized countries, oral health care based on private or public systems is available to the population, but many developing regions have few oral health services. Periodontitis is a burden on social and individual healthcare budgets (Petersen and Ogawa, 2005). According to the Korean National Health Insurance Service (NHIS), it is one of the most common oral diseases in Korea. The cost of health insurance for periodontitis represents a large and continuously growing proportion of NHIS expenditure (2013 Korean statistical annual report, http://www.nhis.or.kr). Thus, the need for risk factor-based periodontitis preventive strategies has increased.

Periodontitis is an inflammatory disease characterized by the host-mediated destruction of soft tissue caused by many factors, including stimulation of osteoclastogenesis (Marchbanks and Campbell, 1976). Various immune response genes are differentially expressed in this disease (Ohlrich et al., 2009). Moreover, it has recently been shown that polymorphisms of many of these, including single nucleotide polymorphisms (SNPs) in IL1B, IL6, TGF-β1, and IL8, amongst others, are associated with chronic periodontitis (Ohlrich et al., 2009; Chen et al., 2014; Gabriela Teixeira et al., 2014; Huang et al., 2015; Lavu et al., 2015).

Caspases are intracellular proteases involved in apoptosis-, pyroptosis-, necroptosis- and autophagy-mediated cell death (Los et al., 1999; Shalini et al., 2015). Many previous studies have elucidated the relationship between caspases and periodontitis. For instance, *Aggregatibacter actinomycetemcomitans* infection enhances apoptosis through a caspase 3-dependent mechanism (Kang et al., 2012). Furthermore, caspase 3 expression increases with progression of gingival inflammation, probing depth, and clinical attachment level in periodontitis (Pradeep et al., 2014), and tissue destruction by *Porphyromonas gingivalis* involves both caspase 1-dependent and -independent mechanisms (Taxman et al., 2006). Activation of caspases 3 and 7 is considerably enhanced in gingival tissue from periodontitis patients (Bantel et al., 2005), in whose osteoblasts the activation of caspase 8 has also been observed (Mori et al., 2009).

In the present study, we investigated whether the CASP gene polymorphisms are associated with periodontal disease risk in the Korean population.
MATERIAL AND METHODS

Participants

This study was approved by the Institutional Review Board of the College of Dentistry, Dankook University, Cheonan, Republic of Korea (IRB No. H-1204/004/001). Written consent forms were obtained from all subjects. To investigate the association between CASP genes and chronic periodontitis susceptibility, 51 chronic periodontitis patients and 33 controls lacking the disease were recruited. Characteristics of the study groups are shown in Table 1. All individuals received oral and dental examinations. The periodontitis diagnosis criteria used are cited in Kang et al. (2015).

Table 1. Characteristics of the subjects included in this study.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Controls</th>
<th>Periodontitis patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of subjects</td>
<td>33</td>
<td>51</td>
</tr>
<tr>
<td>Male</td>
<td>24</td>
<td>25</td>
</tr>
<tr>
<td>Female</td>
<td>9</td>
<td>26</td>
</tr>
<tr>
<td>Age (mean ± SD)</td>
<td>24.9 ± 3.2</td>
<td>49.68 ± 9.9</td>
</tr>
</tbody>
</table>

SD = standard deviation.

DNA extraction

Genomic DNA was prepared from buccal cells. Before collecting cells from the inside of a patient’s mouth, we confirmed the absence of any substance that might interfere with our DNA typing method by requiring patients to wash their mouths with water. Two sterilized swabs were used, one for each cheek, to collect cells from the buccal region. A swab was placed on the side of the cheek, and scraped across it over 10 times to obtain a sufficient quantity of epithelial cells. Each sample was extracted according to the DNA isolation kit manufacturer protocol (Roche, Mannheim, Germany).

SNPs selection and genotyping

We first searched for CASP genes containing SNPs (CASP1 to CASP10, CASP12, CASP14, and CASP8AP2) in the National Centre for Biotechnology Information (NCBI) database (http://www.ncbi.nlm.nih.gov/gene/). The selection criteria for exonic, promoter, and intronic SNPs in each gene were as follows: SNPs had to have 1) a minor allele frequency >10%; 2) heterozygosity >0.1; 3) known genotype frequencies in Asian populations; and 4) been examined in a previous study. We finally selected 201 CASP gene SNPs to investigate their association with chronic periodontitis (Table S1). Genotyping of all SNPs was carried out using an Axiom™ Genome-Wide Human Assay (Affymetrix, Inc., Santa Clara, CA, USA). All of the experimental procedures involved in this assay were performed at the Theragen Etex Corporation (Suwon, Republic of Korea) following the manufacturer protocol.

Statistical analysis

Hardy-Weinberg equilibrium (HWE) was assessed in both the patient and control groups using SNPStats (http://bioinfo.iconcologia.net/index.php?module=Snpsstats). Genotype models (codominant, dominant, recessive, and log-additive) and logistic regression analysis were applied
to determine whether differences in genotype frequencies between the periodontal disease and control groups were significant (Kim et al., 2013; Seok et al., 2014). SNPStats and SPSS 18.0 (SPSS, Inc., Chicago, IL, USA) were used for the statistical analysis. Logistic regression was conducted to evaluate odds ratios (ORs), 95% confidence intervals (95%CIs), and P values. Linkage disequilibrium (LD) blocks and haplotypes were identified using Haploview version 4.2 (Barrett et al., 2005). For the statistical tests, the significance level for P values was set at 0.05.

RESULTS

A total of 201 CASP gene SNPs were analyzed (Table S2). Of these, only variants of CASP3 showed a significant association with periodontitis (P < 0.05). Table 2 shows the genotypic and allelic frequencies of these CASP3 polymorphisms in each study group [rs12108497, -894T>C; rs46476002, intron; and rs113420705, 5'-untranslated region (UTR)].

<table>
<thead>
<tr>
<th>SNP</th>
<th>Genotype or allele</th>
<th>Control [N (%)]</th>
<th>Periodontitis [N (%)]</th>
<th>Model</th>
<th>OR (95%CI)</th>
<th>P</th>
<th>Fisher exact P</th>
</tr>
</thead>
<tbody>
<tr>
<td>CASP3</td>
<td>T/T</td>
<td>18 (60.0)</td>
<td>13 (25.5)</td>
<td>Codominant 1</td>
<td>3.52 (1.30-9.56)</td>
<td>0.013</td>
<td></td>
</tr>
<tr>
<td>rs12108497</td>
<td>-894T&gt;C</td>
<td>11 (36.7)</td>
<td>28 (54.9)</td>
<td>Codominant 2</td>
<td>13.65 (1.57-121.94)</td>
<td>0.018</td>
<td>0.006</td>
</tr>
<tr>
<td></td>
<td>C/C</td>
<td>1 (3.3)</td>
<td>10 (19.6)</td>
<td>Dominant</td>
<td>4.38 (1.67-11.50)</td>
<td>0.002</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Recessive</td>
<td>7.07 (0.86-58.31)</td>
<td>0.024</td>
<td>0.05</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Log additive</td>
<td>3.61 (1.61-8.12)</td>
<td>0.0007</td>
<td></td>
</tr>
<tr>
<td>rs12108497</td>
<td>C/C</td>
<td>1 (3.3)</td>
<td>10 (19.6)</td>
<td>Dominant</td>
<td>4.38 (1.67-11.50)</td>
<td>0.002</td>
<td></td>
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<td>7.07 (0.86-58.31)</td>
<td>0.024</td>
<td>0.05</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Log additive</td>
<td>3.61 (1.61-8.12)</td>
<td>0.0007</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>3.21 (1.61-8.12)</td>
<td>0.002</td>
<td></td>
</tr>
<tr>
<td>CASP3</td>
<td>T/T</td>
<td>14 (42.4)</td>
<td>7 (14.0)</td>
<td>Codominant 1</td>
<td>3.41 (1.15-10.12)</td>
<td>0.027</td>
<td></td>
</tr>
<tr>
<td>rs46476002</td>
<td>T/G</td>
<td>17 (51.5)</td>
<td>29 (58.0)</td>
<td>Codominant 2</td>
<td>14.00 (2.46-79.55)</td>
<td>0.003</td>
<td>0.002</td>
</tr>
<tr>
<td></td>
<td>Intron G/G</td>
<td>2 (6.1)</td>
<td>14 (28.0)</td>
<td>Dominant</td>
<td>4.53 (1.57-13.01)</td>
<td>0.0037</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Recessive</td>
<td>6.03 (1.27-28.61)</td>
<td>0.0082</td>
<td>0.021</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Log additive</td>
<td>3.64 (1.64-8.08)</td>
<td>0.0005</td>
<td></td>
</tr>
<tr>
<td>rs113420705</td>
<td>C/C</td>
<td>16 (50.0)</td>
<td>10 (19.6)</td>
<td>Codominant 1</td>
<td>3.52 (1.30-9.55)</td>
<td>0.013</td>
<td></td>
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<tr>
<td></td>
<td>5’UTR T/T</td>
<td>15 (46.9)</td>
<td>33 (64.7)</td>
<td>Codominant 2</td>
<td>12.80 (1.38-118.32)</td>
<td>0.025</td>
<td>0.018</td>
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<tr>
<td></td>
<td>C/T</td>
<td>1 (3.1)</td>
<td>8 (15.7)</td>
<td>Dominant</td>
<td>4.10 (1.54-10.91)</td>
<td>0.0038</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Recessive</td>
<td>5.77 (0.69-48.51)</td>
<td>0.05</td>
<td>0.14</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Log additive</td>
<td>3.54 (1.53-8.20)</td>
<td>0.0015</td>
<td></td>
</tr>
<tr>
<td>rs113420705</td>
<td>C/T</td>
<td>17 (52.0)</td>
<td>53 (52.0)</td>
<td>Dominant</td>
<td>3.61 (1.61-8.12)</td>
<td>0.007</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>P = 0.007</td>
<td>2.56 (1.30-5.03)</td>
<td>0.007</td>
<td></td>
</tr>
</tbody>
</table>

SNP = single nucleotide polymorphism; OR = odds ratio; CI = confidence interval; UTR = untranslated region. Bold numbers indicate significant associations.

In the control group, the distribution of rs12108497 genotypes was in HWE (P = 1.00). Frequencies of T/T, T/C, and C/C genotypes were 25.5, 54.9, and 19.6% in the periodontal disease group, and 60.0, 36.7, and 3.3% in the control group, respectively (Table 2). Logistic regression analyses for periodontitis patients and control subjects were carried out. Table 2 shows a significant difference in rs12108497 genotype frequencies (P < 0.05), and demonstrates that this SNP was significantly associated with periodontitis (OR = 3.52, 95%CI = 1.30-9.56, P = 0.013 under codominant model 1; OR = 13.85, 95%CI = 1.57-121.94, P = 0.018 under codominant model 2; OR = 4.38, 95%CI = 1.67-11.50, P = 0.002 under the dominant model; OR = 3.61, 95%CI = 1.61-8.12, P = 0.007 under the log-additive model). The frequency of the rs12108497 C allele was higher in periodontitis patients (47.1%) than in control subjects (21.7%). Moreover, this difference was
The distribution of rs4647602 genotypes in the control group was in HWE (P = 0.43). T/T, T/G, and G/G genotype frequencies were 14.0, 58.0, and 28.0% in the periodontal disease group, and 42.4, 51.5, and 6.1% in the control group, respectively (Table 2). Logistic regression analyses for periodontitis patients and control subjects were carried out. Table 2 indicates that under all genotype models, rs4647602 showed significant P values (<0.05), revealing that this SNP was significantly associated with periodontitis (OR = 3.41, 95%CI = 1.15-10.12, P = 0.027 under codominant model 1; OR = 14.00, 95%CI = 2.46-79.55, P = 0.003 under codominant model 2; OR = 4.53, 95%CI = 1.57-13.01, P = 0.0037 under the dominant model; OR = 6.03, 95%CI = 1.27-28.61, P = 0.0082 under the recessive model; OR = 3.64, 95%CI = 1.64-8.08, P = 0.005 under the log-additive model). The frequency of the rs4647602 G allele was higher in the periodontitis group (57.0%) than in the controls (31.8%), and this difference was found to be statistically significant (OR = 2.84, 95%CI = 1.48-5.45, P = 0.002).

In the control group, the distribution of rs113420705 genotypes was in HWE (P = 0.43). The frequencies of C/C, C/T, and T/T genotypes were 19.6, 64.7, and 15.7% in the periodontitis group, and 50.0, 46.9, and 3.1% in the control group, respectively (Table 2). Logistic regression analyses for each study group were carried out. In Table 2, it can be seen that under each genotype model except for recessive model, the rs113420705 SNP showed significant P values (<0.05), demonstrating an association with periodontitis (OR = 3.52, 95%CI = 1.30-9.55, P = 0.013 under codominant model 1; OR = 12.80, 95%CI = 1.38-118.32, P = 0.025 under codominant model 2; OR = 4.10, 95%CI = 1.54-10.91, P = 0.0038 under the dominant model; OR = 3.54, 95%CI = 1.53-8.20, P = 0.0015 under the log-additive model). The rs113420705 T allele frequency was higher in the periodontitis group (48.0%) than in the control group (26.6%), and this difference was statistically significant (OR = 2.56, 95%CI = 1.30-5.03, P = 0.007).

These results indicate that the C, G, and T alleles of rs12108497, rs4647602, and rs113420705, respectively, may constitute risk factors for the development of periodontitis.

In order to further investigate the relationship between these three CASP3 SNPs and periodontitis, the presence of LD blocks between rs12108497, rs4647602, and rs113420705 was assessed using Haploview version 4.2. One LD block was constructed (rs4647602 and rs113420705; D’ = 0.968, r^2 = 0.69; Figure 1), defining three haplotypes. Of these haplotypes, two (TC and GT) were found to be associated with periodontitis (TC, P = 0.0039; GT, P = 0.002; Table 3).

![Figure 1. Linkage disequilibrium block defined by the CASP3 single nucleotide polymorphisms tested.](image-url)
DISCUSSION

Chronic periodontitis could represent a persistent reservoir of microparticles released by multiple cells in an inflammatory context, and hence might be associated with increased prevalence of systemic diseases (Badran et al., 2015). The link between periodontitis and diabetes mellitus is well known (Grossi and Genco, 1998). Furthermore, the prevalence and incidence of coronary heart disease (Bahekar et al., 2007) and the risk of atherosclerotic cardiovascular disease (Dietrich et al., 2013) is increased in periodontitis. Periodontal treatment reduces the risk of cardiovascular disease by lowering plasma levels of inflammatory and thrombotic markers and adhesion molecules (Saffi et al., 2015). Periodontitis is related to rheumatoid arthritis, and treatment for the former has an effect on both the latter and systemic inflammation (Payne et al., 2015). In addition, preeclampsia (Ha et al., 2014), other complications during pregnancy (Zi et al., 2015), and inflammatory bowel disease (Vavricka et al., 2013), amongst other conditions, have been shown to be associated with chronic periodontitis.

This disease is significantly linked to socioeconomic influences, such as low income, low education level, socioenvironmental factors, race, and ethnic group (Petersen and Ogawa, 2005). Indeed, it is a more prevalent and severe problem in developing countries (Pilot, 1998). In the US, periodontitis prevalence is highest in Hispanics (63.5%) and non-Hispanic blacks (59.1%), followed by non-Hispanic Asian Americans (50.0%), and is lowest in non-Hispanic whites (40.8%; Eke et al., 2015). In groups of similar socioeconomic status, access to social security and possession of health insurance has been shown to be important risk factors for periodontitis (Singh et al., 2014). This disease is a major public health problem worldwide and effective prevention methods are needed.

Periodontitis risk is influenced by many factors, and several reports have shown that genetic characteristics make an important contribution. A study of self-reported periodontal disease in a Virginia twin population showed the key role of genetic factors for adult-onset periodontal disease risk (Corey et al., 1993), and oral morphology and function have been observed to be influenced by genetic components (Kurushima et al., 2015). Furthermore, data from twin studies indicate that both genetic and environmental circumstances affect periodontal disease (Michalowicz, 1994). Recently, many investigations have reported associations between individual genetic polymorphisms and chronic periodontitis susceptibility, involving many genes including \(\text{IL1B}, \text{IL1RN}, \text{TNF-}\alpha, \text{TGF-}\beta1, \text{IL8}, \text{CCL5}, \text{COX2}, \text{DEFB1}, \text{MMP1}, \text{MMP9}, \text{STAT1}, \text{CD14}, \text{IKKI}, \text{LT-}\alpha, \text{MiR146a}, \text{MiR499}, \text{OPG}, \text{RANKL}, \text{and VDR}.

In this study, we investigated \textit{CASP} genes in the NCBI database containing SNPs (\textit{CASP1} to \textit{CASP10}, \textit{CASP12}, \textit{CASP14}, and \textit{CASP8AP2}). Caspases are proteolytic enzymes known for their role in controlling cell death and inflammation. However, their functions extend beyond this to include cell proliferation, tumor suppression, differentiation, neural development, and axon guidance and aging (Shalini et al., 2015). Our results revealed a relationship between \textit{CASP3}
SNPs and periodontitis. A number of previous enquiries have established an association between caspases and chronic periodontitis but none have reported a link between this condition and CASP gene polymorphism. For example, the concentration of caspase 3 in gingival crevicular fluid and serum has been seen to proportionally increase with the progression of periodontal disease (Abuhussein et al., 2014; Pradeep et al., 2014). Moreover, *Aggregatibacter actinomycetemcomitans* infection enhances apoptosis of osteoblastic cells, reducing osseous coupling through a caspase 3-dependent mechanism (Kang et al., 2012; Pacios et al., 2013). Caspase 3 is also involved in the *Porphyromonas gingivalis*-derived lipopolysaccharide-mediated activation of the MAPK signaling pathway in human periodontal ligament fibroblasts (Seo et al., 2012). In addition to its role in apoptosis, caspase 3 is crucial for bone formation and the differentiation of bone marrow stromal stem cells into osteoblasts, due to its influence on the TGF-β/SMAD2 pathway and cell cycle progression (Miura et al., 2004).

In the current work, the C, G, and T alleles of rs12108497, rs4647602, and rs113420705, respectively, were found to be associated with the development of periodontitis. rs12108497 has previously been linked to stomach and gastric cancers, while rs4647602 is associated with squamous cell carcinoma of the head and neck and atrial fibrillation, and rs113420705 is involved in coronary artery lesion formation in Kawasaki disease. Periodontitis leads to inflammatory responses in the atrial myocardium, disturbing the structural properties and electrophysiology of the atrium, facilitating its fibrillation (Yu et al., 2010). Moreover, periodontal bone loss is associated with complex multiple coronary lesions (Romagna et al., 2012). Altogether, these results might provide further evidence for the role of caspase 3 in chronic periodontitis events, such as cell apoptosis, inflammation, and bone loss.

The prevalence of edentulousness in the elderly is low in many industrialized countries but remains high in developing regions. Periodontitis and tooth loss are linked to quality of life and chronic diseases, and thus constitute a global burden. Early diagnosis utilizing genetic factors and prevention of chronic periodontitis could improve oral health and reduce social and individual healthcare costs.

To conclude, CASP3 gene polymorphisms may be markers of susceptibility to periodontal disease in the Korean population. Additional studies with larger sample sizes and a focus on different populations will need to be conducted to confirm our results.

Conflicts of interest

The authors declare no conflict of interest.

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Supplementary material

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